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(54) Title: ALGINATE CONTAINING ANTIMICROBIAL COMPOSITION

(57) Abstract

There is provided a composition comprising an admixture of a finely divided alginate (or a salt or derivative thereof) together with a finely divided carrier material. The composition overcomes the problems associated with applying gel-forming alginates to a body surface without formation of a clumpy paste that leads to local irritation. An admixture of sodium alginate and a water-soluble glass carrier material is preferred. Optionally, the alginate and carrier material each have a particle size of less than 150 μ m diameter and are present in a weight ratio of 20:80 to 80:20. The presence of the carrier aids even gel formation and also promotes wound healing.

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1 ALGINATE CONTAINING ANTIMICROBIAL COMPOSITION 2 The present invention relates to an anti-microbial 3 composition for use in medical or veterinary 5 applications. A wide variety of gels, creams, ointments, lotions etc 7 are available for application to a body surface. 8 exact content of such compositions generally depends 9 upon the purpose of application which may be, for 10 example, to clean a body surface, to promote healing of 11 any wound or injury, to prevent an exposed area of the 12 body from drying out, to prevent infection etc. 13 certain circumstances the composition may include an 14 15 active ingredient which is administered to the patient 16 by application of the composition. 17 18 One example of a commercially available gel is INTRASITE™ produced by Smith & Nephew Ltd. 19 hydrogel contains hydrated carboxymethylcellulose as 20 its main ingredient, and is applied to wounds in gel 21 form as a primary treatment in order to clean the 22 exposed surface by aiding removal of cell debris, dirt 23 In addition to acting as a sloughing agent, the 24 gel also keeps the wound from drying out, thereby 25

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promoting healing. 1 2 Another example of a gel suitable for use on a wound 3 dressing is described in EP-A-0586260 of Courtaulds 4 Fibres Ltd. The gel disclosed is an alginate gel 5 having an alginate content of 2 to 11 percent by 6 7 weight. 8 Surgical dressings based on gel forming alginates have 9 a significant contribution to make in wound management 10 and are generally presented as preformed components of 11 gels and pastes and as fibres of calcium or mixed 12 13 calcium/sodium salts. 14 In alginate-based surgical dressings the starting raw 15 material is usually the sodium salt which is supplied 16 by the alginate producer as a dry powder. Attempts to 17 utilise alginate as topical powders for direct 18 application to wounds have not proved successful. This 19 is because the irregularly dispersed powder does not 20 wet easily and clumping occurs leading to clusters of 21 dry particles which can be sites of local irritation. 22 There is incomplete gelling as a result and the desired 23 sealing of the wound with a smooth hydrogel coating is 24 not achieved. 25 26 It has now been found that an admixture of finely 27 divided alginate (the term "alginate" being used herein 28 to refer to alginates, the derivatives and salts 29. thereof) and a different finely divided carrier 30 material can be applied to wounds or other moist body 31 surfaces. The combination of the carrier material 32 together with the alginate facilities the formation of 33 an even gel coating and the avoidance of clumping. 34

Suitable carrier materials include proteins (eg

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casein), salts (eg sodium, zinc, calcium, magnesium and 1 potassium salts) and water-soluble glass. Desirably 2 3 the carrier material is water-soluble or water 4 miscible. 5 6 More surprisingly, it has been found that the alginate/carrier combination acts in synergy to promote 7 healing and cell growth. For example, in animal 8 implant studies which compared alginate powder alone 9 and a water-soluble glass powder alone with a blend of 10 both, it was demonstrated that tissue response was 11 clearly better for the mixed powders than that seen 12 13 with either material on its own. In particular at 14 days after implantation there was little evidence of 14 the inflammatory cells which were residually present in 15 the single material implant sites. 16 17 Viewed from one aspect the present invention provides 18 an admixture of alginate or a derivative or salt 19 20 thereof together with a carrier material. Generally both main components are finely divided, i.e. are in 21 22 powder, particulate or granular form. 23 Desirably the finely divided alginate and carrier 24 material components may each have a diameter size of 25 150µm or less. Preferably the mode particle size for 26 either component is 100 µm or less. More preferably the 27 mode particle size for either component is $60\mu\text{m}$ or 28 29 less, for example $30-60\mu m$. 30 The two components may be combined together in any 31 suitable mixture. Suitable mixtures include those 32 having a ratio of from 20:80 to 80:20 (% by weight) of 33 alginate:carrier. Preferred mixtures include those 34 having an alginate:carrier ratio in the range of 20:80 35 to 50:50, preferably 20:80 to 30:70, for example 25:75. 36

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Water-soluble glasses are a preferred form of carrier 1 material. The use of glasses which can dissolve in 2 water and body fluid and which are applied internally 3 of the body are well-known. These glasses are formed 4 from phosphorus pentoxide and may be modified to 5 dissolve over a period of minutes, months or even 6 years, as required. To date, such glasses have been 7 used, in medicine, for the controlled release of a 8 number of agents, for example, drugs, hormones and 9 trace elements, but in each case the glass has been 10 applied internally of the body to allow the agent to 11 leach out into the body's circulatory system. 12 13 It is known that certain glasses, in which the usual 14 glass former, silicon dioxide, of traditional glasses 15 is replaced with phosphorus pentoxide as the glass 16 former, are soluble in water and body fluids. 17 of dissolution is controlled largely by the addition of 18 glass modifiers such as calcium and magnesium oxide. 19 In simple terms, the greater the concentration of the 20 modifier the slower is the rate of dissolution. 21 rates of dissolution which can be imparted to the 22 glasses may range from minutes to months or even to 23 several years. It is known to include in such 24 compositions quantities of trace elements such as 25 copper, cobalt and selenium which will be released from 26 the glass as it slowly dissolves over the selected 27 period of time. 28 29 The use of water-soluble glasses has been described for 30 a variety of purposes in the literature. For example, 31 UK Patent Specifications numbers 1,565,906, 2,079,152, 32 2,077,585 and 2,146,531 describe the gradual 33 dissolution of the glasses as providing a means of 34 controlled release of drugs, hormones, fungicides, 35 insecticides, spermicides and other agents with which 36

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the glasses have been impregnated. The glasses are 1 used for example in the form of an implant or bolus. 2 3 UK Patent Specification number 2,030,559 describes the 4 use of selenium-impregnated water-soluble glass for 5 providing controlled release of the selenium as a trace 6 element into cattle and sheep, the glass being applied 7 as a subcutaneous insert. UK Patent Specification 8 number 2,037,735 also describes a subcutaneous implant 9 of water-soluble glass, and in this case the glass is 10 impregnated with copper; minor quantities of trace 11 elements such as boron, arsenic, iodine, manganese, 12 chromium, silver, gold and gallium may also be 13 14 included. 15 Water-soluble glass has also been proposed for use in 16 prosthetics, for example in UK Patent Specification 17 number 2,099,702, and for use in anticorrosive paints, 18 19 as described in UK Patent Specification number 2,062,612. Further the literature provides for the use 20 21 of such glasses in the controlled release of ferrous 22 and ferric ions into the human or animal body by 23 ingestion or implantation of the glass (UK Patent Specification number 2,081,703), and for the use of 24 glasses in the controlled release of ions such as 25 lithium, sodium, potassium, caesium, rubidium, 26 polyphosphate, calcium and aluminium to patients by 27 inclusion of the glass in a drip feed line (UK Patent 28

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WO-A-89/01793 relates to apparatus for antimicrobial use in passage of fluid to or from a living body, the apparatus comprising a conduit for insertion into the body, a reservoir for fluid and a connector member for connecting said conduit to said reservoir external of the body, wherein said connector member includes a

Specification number 2,057,420).

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water-soluble glass impregnated with elemental silver 1 or a compound of silver, said water-soluble glass 2 defining at least a part of a passageway for fluid to 3 flow between the reservoir and the conduit. 4 Desirably the water-soluble glass is a silver 6 containing water-soluble glass. Advantageously the 7 silver content will be introduced into the glass 8 9 composition in the form of silver orthophosphate. 10 Suitable glasses include, for example, the ARGLAES™ 11 glass of Giltech Limited. 12 13 Preferably, said glass is adapted by the use of glass 14 modifiers to give a sustained release of silver ions 15 over a set period. 16 17 In one embodiment the water-soluble glass comprises an 18 alkali metal oxide M20, an alkaline earth oxide M0, 19 phosphorus pentoxide P₂O₅ and silver oxide (Ag₂O) or 20 silver orthophosphate (Ag₃PO₄). 21 22 Most preferably, said glass contains not more than 40 23 mole % M_2O or MO, not less than 10 mole % M_2O or MO, and 24 not more than 50 mole % nor less than 38 mole % 25 phosphorus pentoxide, with the inclusion of 0.05 to 5.0 26 mole % silver oxide or orthophosphate. 27 28 Said alkali metal oxide may be sodium oxide (Na20), 29 potassium (K20) or a mixture thereof; and said alkaline 30 earth oxide may be calcium oxide (CaO), magnesium oxide 31 (Mg0), zinc oxide (Zn0) or a mixture thereof. 32 33 The glass may also contain less than 5 mole % silicon 34 dioxide (SiO₂), boric oxide (B₂O₃), sulphate ion (SO₄²⁻), 35 a halide ion, copper oxide (CuO) or a mixture thereof. 36

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Typically the soluble glasses used in this invention 1 comprise phosphorus pentoxide (P2O5) as the principal 2 3 glass-former, together with any one or more glass-modifying non-toxic materials such as sodium 4 oxide (Na_20), potassium oxide (K_20), magnesium oxide 5 (Mg0), zinc oxide (Zn0) and calcium oxide (Ca0). rate at which the silver-release glass dissolves in 7 fluids is determined by the glass composition, В 9 generally by the ratio of glass-modifier to glass-former and by the relative proportions of the 10 glass-modifiers in the glass. By suitable adjustment 11 of the glass composition, the dissolution rates in 12 water at 38°C ranging from substantially zero to 13 25mg/cm²/hour or more can be designed. However, the 14 most desirable dissolution rate R of the glass is 15 between 0.01 and 2.0mg/cm²/hour. The water-soluble 16 glass is preferably a phosphate glass, and the silver 17 may advantageously be introduced during manufacture as 18 silver orthophosphate (Ag₃PO₄). 19 The content of silver and other constituents in the glass can vary in 20 accordance with conditions of use and desired rates of 21 release, the content of silver generally being up to 5 22 mole %. While we are following convention in 23 24 describing the composition of the glass in terms of the mole % of oxides, of halides and of sulphate ions, this 25 is not intended to imply that such chemical species are 26 present in the glass nor that they are used for the 27 batch for the preparation of the glass. 28

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The optimum rate of release of silver ions into an aqueous environment may be selected by circumstances and particularly by the specific function of the released silver. The invention provides a means of delivering silver ions to an aqueous medium at a rate which will maintain a concentration of silver ions in said aqueous medium of not less than 0.01 parts per

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million and not greater than 10 parts per million. 1 some cases, the required rate of release may be such 2 that all of the silver added to the system is released 3 in a short period of hours or days and in other 4 applications it may be that the total silver be 5 released slowly at a substantially uniform rate over a 6 period extending to months or even years. 7 particular cases there may be additional requirements, 8 for example it may be desirable that no residue remains 9 after the source of the silver ions is exhausted or, in 10 other cases, where the silver is made available it will 11 be desirable that any materials, other than the silver 12 itself, which are simultaneously released should be 13 physiologically harmless. In yet other cases, it may 14 be necessary to ensure that the pH of the resulting 15 solution does not fall outside defined limits. 16 17 The glass may be formed by a number of methods. 18 simply be cast by conventional or centrifugal 19 procedures, or it may be prepared via one or more 20 stages of rod, fibre or tube drawing. Other 21 preparation techniques include foamed glass. 22 glass formation it will be comminuted into finely 23 24 divided form. 25 With regard to the alginate component, derivatives and 26 salts of alginates are acceptable for use in the 27 Sodium and calcium salts of present invention. 28 alginate or a combination of these two salts is 29 preferred. Sodium alginate is especially preferred. 30 31 In one preferred embodiment, the composition of the 32 present invention is an admixture of sodium alginate 33 powder and water soluble glass (eg ARGLAES™ of Giltech 34 Limited) in a ratio of alginate: glass of 25:75 by 35 weight. Preferably, the water soluble glass releases 36

calcium ions as it dissolves. The calcium ions
displace some of the sodium ions in the sodium alginate
thus forming calcium alginate. The presence of calcium
alginate stabilises the alginate gel.

The composition may be pre-mixed, or alternatively the

The composition may be pre-mixed, or alternatively the alginate may be kept separately from the carrier material and the ingredients admixed together immediately prior to use. This enables a particular blend to be formulated to suit the wound or condition in question.

Optionally, the composition of the present invention may contain an active ingredient. The term "active ingredient" is used herein to refer to any agent which affects the metabolism or any metabolic or cellular process of the patient (including growth factors and living cells), promotes healing, combats infection, hypergranulation or inflammation. Antibiotics and other anti-bacterial agents, steroids, painkillers etc are all suitable. Optionally, the active ingredient may be in delay-release or controlled-release form.

The composition of the present invention may be used to clean a body surface, to promote healing of a wound or injury, to prevent an exposed area of the body from drying out or to prevent infection.

 In a further aspect the present invention provides a method of treating the human or non-human (preferably mammalian) animal body, said method comprising applying a finely divided admixture of an alginate (a derivative or salt thereof) and a carrier material, such as a (preferably silver-containing) water-soluble glass, to a body surface, for example to a wound.

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1 The invention will now be further described with 2 reference to the figures: 3 4 Fig 1 illustrates a mass of inflammatory cells at the site of implantation of a composition of just silver 5 6 ion releasing glass, 7 days after implantation. 7 8 Fig 2 illustrates a mass of inflammatory cells and the 9 damage to the muscle bed at the site of implantation of alginate, 2 days after implantation. 10 11 12 Fig 3 is a higher magnification of the same tissue 13 block as in Fig 2. 14 15 Fig 4 illustrates a mass of inflammatory cells sitting on and infiltrating the muscle bed at the site of 16 17 implantation of a composition of just alginate, 7 days 18 after implantation. 19 20 Fig 5 is a higher magnification of the same tissue 21 block as in Fig 4. 22 23 Fig 6 illustrates a number of inflammatory cells and 24 the broken up muscle bed at the site of implantation of 25 a composition of alginate and a water soluble glass 26 carrier, 2 days after implantation. 27 28 Fig 7 illustrates a number of inflammatory cells and a normal muscle bed at the site of implantation of a 29 30 composition of alginate and a water soluble glass 31 carrier, 7 days after implantation. 32 33 and with reference to the following, non-limiting, 34 examples.

1 EXAMPLE 1 2 To determine the tissue response to the powdered 3 biomaterials using a rat model and further to determine 4 whether combining the two materials had a significant 5 effect on the response. 6 7 8 <u>Materials</u> 9 CRG/silver powder [D301893 Ag 3 mole%] .. 10 CRG/Aq Alginate powder [lot No 544831] 11 Alginate 12 CRG/silver powder and Alginate powder [50:50] mix Alginate/CRG/Ag 13 14 The silver containing controlled release glass (herein 15 referred to as "CRG/silver") had the following 16 17 composition Na₂C 27.5 mole % 18 Cal 22 mole % 19 Ag_20 3.5 mole % 20 $P_{2}O_{5}$ 47 mole % 21 The silver content of the glass was added in the form 22 of silver orthophosphate, but is expressed as "silver 23 oxide" according to convention. 100% of the glass 24 25 particles had a diameter of less than 53 µm. 26 The alginate used was a pure sodium alginate salt, 27 28 commercially available as $Manucol^{re}$ LKX of Kelco International Limited, United Kingdom. The volume mode 29 particle size of the sodium alginate is 41.46 µm and 30 99.4% of the particles had a diameter of less than 31 32 49.99µm. 33 All materials were supplied in powder form. 34 Alginate/Ag mix was prepared by hand. The materials 35 were not sterilised before implantation. No infection 36

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problems were encountered during the procedures. 1 2 3 Method 4 Adult, black and white hooded rats of the Lister strain 5 (approximately 200g) were used for all procedures. 6 7 Appropriate surgical methods were employed by experienced personnel, and all procedures were carried 8 out as detailed in UK Home Office licence No 9 PP140/01099. 10 11 A small incision was made above the lumbar sacral 12 vertebrae, and the muscle bed on either side of this 13 incision was exposed by blunt dissection. A pocket was 14 created in the muscle fibres and approximately 2mg of 15 the powdered material was carefully placed into this 16 pocket. Inevitably, some powder material was deposited 17 on the muscle bed surface and contacted subcutaneous 18 tissue. Animals were sacrificed at 2, 7 and 14 days 19 20 using a schedule one method. 21 22 Following sacrifice, the tissue was examined for any obvious signs of inflammation, and a block of 23 tissue/muscle containing the implant site was removed. 24 The block was immediately frozen, sectioned on a 25 cryostat microtome to produce sections 7µm thin and 26 stained using haematoxylin and eosin. 27 The sections were examined by light microscopy. 28 29 30 Results 31 32 CRG/Aq 33 2 days 34 35

There were no signs of gross inflammation when the

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animals were sacrificed. Following staining the site 1 2 could be seen to be heavily inflamed. The muscle was widely infiltrated with neutrophils, and the muscle 3 fibres were disrupted. A black particulate material 4 5 (believed to be an Ag/Ag complex) was evident and neutrophils were very densely packed around these 6 7 particles. 8 9 7 days 10 Although the muscle site appeared clean, there was a 11 large volume of clear exudate present at each implant 12 13 The exudate had produced a swelling under the skin at the site of the implantation. Following 14 staining, a mass of inflammatory cells were seen to be 15 16 present at the site (Fig 1). These cells appeared to 17 be predominantly neutrophils. The muscle fibres 18 appeared normal and there was no evidence of necrotic 19 tissue, though there remained some inflammatory infiltration. Particulate matter was present though 20 21 not black in this case. It looked more like a 22 degrading glass. The silver could not be detected at 23 this time. 24 25 14 days 26 27 The exudate and associated swelling had subsided by 28 this time, however when the site was exposed there was evidence of tissue damage (believed to be necrosis) on 29 the muscle bed and in contiguous subcutaneous tissue. 30 Following staining extensive inflammation was apparent, 31 32 and there was evidence of necrotic tissue. However, only a small area was affected. Some dark, particulate 33 34 material was also evident. This may be a silver 35 Degrading glass material is clearly present

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at the site.

1 Alginate

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3 2 days

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No gross signs of inflammation were present when the 5 animals were sacrificed. However, the alginate was clearly visible on and around the implant site as a 7 8 "messy" gel. Following staining, large numbers of 9 inflammatory cells could be seen (Fig 2), the muscle bed was damaged and the muscle fibres were disturbed 10 and infiltrated with these cells. This was possibly 11 12 due to the presence of small particulate material 13 invading the muscle and stimulating an inflammatory response. However, there was no evidence of necrotic 14 15 response.

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Fig 3 shows a higher magnification of the response from the same tissue block as Fig 2. Inflammatory cells can be seen invading the muscle fibres. Most of the pink stained material visible was alginate, clearly well dispersed. Muscle fibres (also stained pink) could be seen in the top right corner. Alginate could be seen, stained pink.

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7 days

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No signs of gross inflammation were evident when the 27 animals were sacrificed. No alginate could be seen at 28 this time, and the muscle bed appeared clean. 29 Following staining (Fig 4), large numbers of 30 inflammatory cells could be seen remaining at the 31 implant site. However, there was very little evidence 32 of alginate remaining at the site even when the site 33 was observed under higher magnification (Fig 5). 34 result was very similar to that observed with the Ag at 35 7 days although in this case there was no exudate 36

15 build-up. 1 2 3 14 days 4 5 No sign of gross inflammation was present when the 6 animal was sacrificed. Following staining, large 7 numbers of inflammatory cells were evident at the 8 implant site. There was some evidence of alginate 9 remaining at the site, but only very little. no evidence of necrosis or damage to the tissue. 10 11 12 Alginate/CRG/Ag 13 14 2 days 15 16 There were no gross signs of inflammation when the 17 animals were sacrificed, and the muscle bed appeared 18 clean. Following staining (Fig 6), the muscle fibres 19 could be seen to be disturbed and the muscle bed to be 20 broken up. This was likely to be due to the 21 particulate matter stimulating infiltration of 22 inflammatory cells. However, there appeared to be 23 fewer inflammatory cells at the implant site or 24 infiltrating the muscle than was evident when the 25 materials were examined alone. There was only little 26 evidence of particulate material remaining at the site. 27 Once again, this appeared to be a degrading glass. 28 29 7 days 30 31 There were no gross signs of inflammation when the animals were sacrificed. Following staining (Fig 7),

animals were sacrificed. Following staining (Fig 7),
large numbers of inflammatory cells could be seen at
the implant site. There was some particulate material
present, though it was not clear what this was. The

response was similar to that seen at 2 days. However,

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the muscle bed now seems normal with the muscle fibres 1 2 intact. The result was very similar to that seen with the materials examined alone at the same time period. 3 14 days There were no signs of gross inflammation at the implant site following sacrifice. Staining showed a 8 9 clean muscle block with only little evidence of inflammatory cells. The response at 14 days with the 10 11 mixed materials, was clearly better than that seen with 12 either material when examined alone. No evidence of any particulate material could be found at this time. 13 14 15 Conclusion 16 The majority of inflammation that is seen with these 17 18 samples can probably be attributed to: 19 20 the surgical procedure itself; we are examining a. the tissue response within the normal wound 21 22 healing time; 23 24 b. the fact that the material has been applied in 25 power/particulate form; this will inevitably lead 26 to a more extensive inflammation. 27 28 Nevertheless, differences have been noted in the 29 responses to the materials examined. Silver containing 30 CRG gave rise to a considerable exudate which was at 31 its most severe, certainly most obvious at 7 days. 32 This exudate was clearly visible under the skin as a 33 lump, and the area was obviously painful to the animal. 34 On sacrifice the exudate was revealed as a clear, subcutaneous fluid. At 14 days the exudate had 35 subsided, although there remained a "sore" on the skin. 36

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1	When exposed, the implant site, particularly the muscle
2	bed surface and the subcutaneous tissue in contact with
3	the implant site, was damaged. Histology showed that
4	there was some evidence of necrotic tissue, though this
5	was minimal.
6	
7	The alginate alone produced a "messy" gel on the muscle
8	surface at 2 days, but subsequent time periods showed a
9	clean muscle bed. Inflammation was associated with the
10	implant site at all time periods. However, there was
11	no evidence of damage or necrotic tissue. Although the
12	alginate is clearly dissolving, traces of alginate
13	could still be found at the site for 14 days.
14	
15	The alginate/silver mix seemed to attract less cells to
16	the site at 2 days. At 7 days the response was fairly
17	similar to that seen with the samples examined alone
18	and no exudate was formed. However, after 14 days the
19	healing response seemed much accelerated with this
20	sample. Clean, normal muscle tissue was observed, with
21	little evidence of inflammatory infiltration.
22	
23	EXAMPLE 2
24	
25	Materials examined:
26	
27	CRG/Ag powder
28	Alginate powder
29	Alginate/CRG/Ag powder
30	
31	All the samples were implanted as powders.
32	
33	Adult, black and white hooded Lister rats
34	(approximately 200g) were used.
35	
36	A small incision was made above the lumber sacral

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18 vertebrae. A pocket was created in the muscle fibres 1 and approximately 5mg of the material was placed into 2 the pocket. The wound was sutured with silk. 3 4 5 Two samples of each material were placed in each animal and two animals used for each time period. 6 were sacrificed at two and seven days. 7 8 At sacrifice the tissue was examined for any obvious 9 signs of inflammation and a block of muscle containing 10 the implant site removed. The block was frozen, 11 sectioned on a microtome at 7 microns and stained by 12 haematoxylin and eosin. 13 14 15 CRG/Ag Powder 16 17 2 days 18 There were no gross signs of inflammation when the 19 animal was sacrificed. Following staining, the site 20 could be seen to be heavily inflamed. The muscle was 21 widely infiltrated with neutrophils, and the muscle 22 fibres disrupted. A black particulate material (Ag/Ag 23 complex) was in evidence and neutrophils were very 24 densely packed around these particles. 25 26 27 7 days 28 Although the muscle site looked clean, there was a 29 large volume of clear exudate present with each animal. 30 The exudate had produced a swelling under the skin at 31 the site of the implant. Following staining, a huge 32 mass of inflammatory cells were present at the implant 33 These cells appear to be predominantly 34

neutrophils. The muscle fibres looked normal, though

there remained a considerably inflammatory cell

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There was some particulate matter 1 infiltration. present, though not black in this case. It looked more 2 like a degrading glass. 3 5 Alginate powder 6 7 2 days 8 9 No gross signs of inflammation when the animal was sacrificed, though the alginate was clearly visible on 10 and around the implant site, as a "messy" gel. 11 12 Following staining, large numbers of inflammatory cells 13 could be seen and the muscle fibres were disturbed and 14 infiltrated with these cells. Alginate could be seen, 15 stained pink. 16 17 7 days 18 19 No gross signs of inflammation when the animal was 20 sacrificed. No sign of alginate at this time. Muscle 21 bed looked very clean. Following staining, large 22 numbers of inflammatory cells could be seen remaining 23 at the implant site, however, there was very little 24 evidence of alginate remaining at the site. 25 was similar to that observed with CRG/Ag at 7 days, 26 although in this case there was no exudate build up. 27 28 Alginate/CRG/Ag 29 30 2 days 31 32 No gross signs of inflammation when the animal was 33 sacrificed. The muscle bed was clean. Following 34 staining, the muscle fibres could be seen to be broken 35 up, however, there were less numbers of inflammatory cells at the implant site or infiltrating the muscle. 36

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There was only little evidence of particulate material 1 2 remaining at the site. Again this looked like a 3 degrading glass. 5 7 days 7 No gross inflammation when the animal was sacrificed. 8 Following staining large numbers of inflammatory cells 9 could be seen at the site of implantation. Again there was some particulate material present (degrading 10 11 glass). The muscle fibres were intact and normal. 12 13 EXAMPLE 3 14 15 Method 16 Other powders have also been combined with alginate to 17 establish whether a) these combinations also formed a 18 gel and b) if any such gel was tacky. 19 20 The powders tried were casein, sodium chloride, zinc 21 oxide, sodium borate, magnesium sulphate, magnesium 22 chloride, calcium tetraborate and potassium iodide. 23 24 Each powder was admixed individually with sodium 25 alginate (Manucol™ LKX) in a ratio of 3:1. The 26 admixture was then applied to a damp simulated wound, 27 covered with a dressing and left for 48 hours. 28 29 Results 30 Admixtures with casein, sodium chloride, magnesium 31 sulphite, magnesium chloride and potassium iodide 32 formed sticky but "lump free" gels. 33 Admixtures with zinc oxide and calcium tetraborate did 34 35 not appear to wet out at all. 36

- 1 The admixture with sodium borate did wet out
- 2 adequately, but formed a rubbery coating on the
- 3 simulated wound which did not stick to the dressing.

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1	CLA	IMS
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3	1.	A composition comprising an admixture of finely
4		divided alginate and a finely divided carrier
5		material.
6		
7	2.	An admixture as claimed in Claim 1, wherein the
8		ratio of alginate:carrier material is in the range
9		20:80 to 80:20 by weight.
10		
11	3.	An admixture as claimed in Claim 2, wherein the
12		ratio of alginate:carrier material is 25:75 by
13		weight.
14		
15	4.	A composition as claimed in any of the preceding
16		Claims, wherein the carrier material is a water
17		soluble glass.
18		
19	5.	A composition as claimed in Claim 4, wherein said
20		water soluble glass releases silver ions during
21		dissolution.
22		
23	6.	A composition as claimed in either one of Claims 4
24		and 5, wherein said water soluble glass releases
25		calcium ions during dissolution.
26		
27	7.	A composition as claimed in any one of the
28		preceding Claims, wherein the alginate is sodium
29		alginate, calcium alginate or a mixture thereof.
30		
31	8.	A composition as claimed in Claim 7, wherein the
32		alginate is sodium alginate.
33		
34	9.	A composition as claimed in any one of the
35		preceding Claims, wherein said finely divided

alginate has a particle diameter of 150 $\mu\mathrm{m}$ or

36

30

prevent infection.

1		less.
2		
3	10.	A composition as claimed in any one of the
4		preceding Claims, wherein said finely divided
5		carrier material has a particle diameter of 150 $\mu \mathrm{m}$
6		or less.
7	·	
8	11.	A composition as claimed in any one of the
9		preceding Claims, wherein said alginate and said
10		carrier material each have a mode particle size of
11		60 μ m or less.
12		
13	12.	A composition as claimed in any one of the
14		preceding Claims, said composition comprising
15		75 parts by weight of a finely divided calcium ion
16		releasing water soluble glass and 25 parts by
17		weight of finely divided sodium alginate, said
18		glass and said alginate each having a mode
19		particle size of 60 μm or less.
20		
21	13.	A method of treatment of a numan or non-human
22		animal body, said method comprising applying to a
23		surface of said body a composition as claimed in
24		any one of Claims 1 to 12.
25		
26	14.	Use of a composition as claimed in any one of
27		Claims 1 to 12 to clean a body surface, to promote
28		healing of a wound or injury, to prevent an
29		exposed area of the body from drying out or to

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Fig. 1

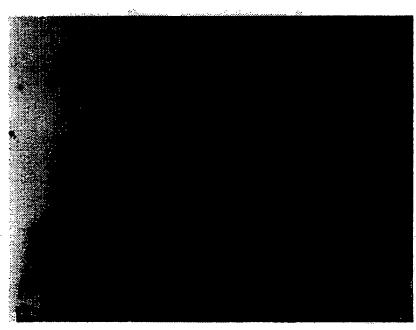


Fig. 2

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Fig. 3

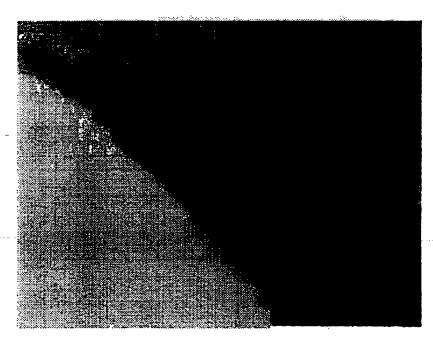


Fig. 4

SUBSTITUTE SHEET (RULE 28)



Fig. 5

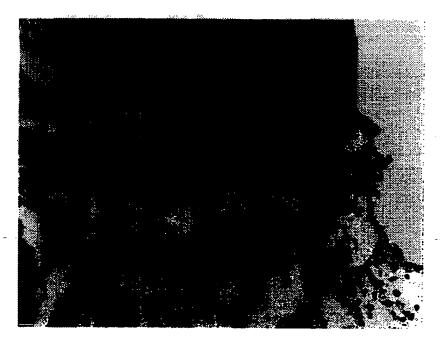


Fig. 6

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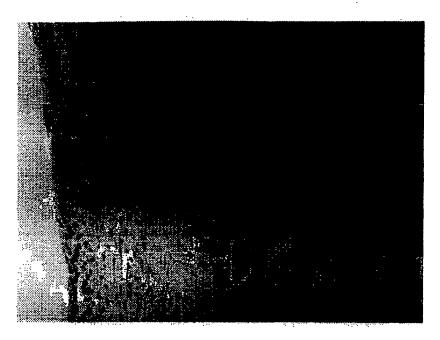


Fig. 7